

Title: "Studies on the role of Rab-GTPase Ypt1p in the nuclear mRNA

surveillance in *Saccharomyces cerevisiae*" [Index No.: 66/17/Life Sc./25]

ABSTRACT

Nuclear degradation of pre-*HAC1* mRNA and subsequent targeting of the resulting precursor mRNA plays a vital role in the activation as well as attenuation of Unfolded Protein Response (UPR) in *Saccharomyces cerevisiae*. Unfolded Protein Response (UPR) is an intracellular signalling pathway that responds to stress causing a burden of unfolded proteins (ER stress) in the ER lumen. To mitigate this situation, baker's yeast *Saccharomyces cerevisiae* produce a huge burst of Hac1p, a transcription factor that activates genes encoding ER chaperones. Hac1p is encoded by *HAC1* pre-mRNA harboring an intron and a bipartite element (BE) at its 3'-UTR, which undergoes a reversible and dynamic intra-nuclear 3'→5' mRNA decay by the nuclear exosome/CTEXT at various phases of UPR. Accelerated decay, in absence of stress leads to a pre-*HAC1* mRNA pool, the majority of which lacking the BE, thereby undergoing inefficient targeting to Ire1p foci consequent to diminished splicing and lower production of mature Hac1p. ER-stress leads to its diminished decay, producing an increased abundance of pre-*HAC1* mRNA population carrying an intact BE leading to its accelerated recruitment to Ire1p foci followed by increased splicing and increased production of mature Hac1p.

Rab-GTPase Ypt1p, which was previously implicated in the ER to Golgi shuttling of proteins was also demonstrated to be involved in the regulation of UPR signalling dynamics by promoting the decay of *HAC1* mRNA. This finding suggests a potential involvement of Ypt1p in the regulatory mechanism for selection and recognition of the *HAC1* pre-mRNA by the nuclear exosome/CTEXT. In this study we show that physical association (as detected by RNA-immunoprecipitation) of Rab-GTPase Ypt1p with *HAC1*-pre mRNA facilitates the recruitment (as detected by ChIP) of NNS/CTEXT/nuclear exosome on to the pre-*HAC1* mRNA and stimulates its nuclear decay in absence of stress. In presence of stress, reduced amount of Ypt1p becomes associated with the pre-*HAC1* thereby lowering the recruitment of exosome/CTEXT/NNS (as detected by Co-immunoprecipitation) on to it, which leads to the production of huge amounts of pre-*HAC1* mRNA with an intact BE. Regulation by Ypt1p relies on its characteristic nuclear localization in absence of ER-stress causing its strong association with pre-*HAC1* mRNA at its 3'-UTR that promotes a sequential recruitment of Nrd1-Nab3p-Sen1p (NNS) complex → CTEXT → nuclear exosome onto the pre-*HAC1* mRNA that eventually stimulates its selective nuclear decay. During induction of UPR, Ypt1p rapidly relocalizes to the cytoplasm with its consequent dissociation from pre-*HAC1* mRNA thereby causing a decreased recruitment of NNS/CTEXT/Nuclear exosome with its consequent diminished 3'→5' degradation by the exosome. This reversible mechanism ensures a timely activation of UPR and its prompt attenuation following the accomplishment of homeostasis. These findings will help to gain an insight into the mechanism of intracellular signalling pathway UPR that is accomplished by differential degradation at the post-transcriptional level.

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